Inter- and Intra-specific variation in egg size among reef fishes across the Isthmus of Panama

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Effects of planktonic food supplies and temperature on pelagic fish larvae are thought to be the primary environmental determinants of adaptive variation in egg size. Differences between the Atlantic and Pacific coasts of Panama in primary production (higher in the Pacific due to upwelling) and temperature (less seasonal in the non-upwelling Caribbean) allow testing such ideas. We compared the volumes, dry weights, and energy content of eggs of 24 species of reef fishes from the two sides of the isthmus during the cool and warm seasons. Both egg volume and egg dry weight were good predictors of egg energy content among species, although not within species. Caribbean species produced larger eggs than their close relatives in the Pacific. In the Pacific, eggs were significantly larger during the cool upwelling season than during the warm, non-upwelling period, with a similar but weaker seasonal pattern evident in the Caribbean. The production of larger eggs in the low-productivity Caribbean is consistent with the hypothesis that species produce larger eggs and offspring when larval food supplies are lower. Parallel patterns of seasonal variation in eggs size and the greater strength of that relationship in the Pacific indicate that temperature drives seasonal variation in egg size within species. The decline in egg size with increasing temperature, a general pattern among ectotherms, may be a physiological side-effect, due to differing effects of temperature on various metabolic processes during oogenesis or on hormones that influence growth and reproduction. Alternatively, the seasonal pattern may be adaptive in these fishes, by affecting larval performance or maintaining a particular timeline of major events during embryonic development.

Keywords: primary productivity, upwelling, tropical reef-fish, egg-size variation, temperature-size rule, life history evolution, Isthmus of Panama

INTRODUCTION

Egg-size is a fundamental life history attribute of marine fishes and invertebrates (Bernardo, 1996; Chambers, 1997; Moran and McAlister, 2009; Collin, 2012). Egg size effects the size, growth rate, and survival of hatchling larvae, and, consequently, has cascading effects on offspring fitness (Allen and Marshall, 2014). A major line of inquiry in evolutionary ecology is to understand intra- and inter-specific variation in maternal investment in egg size and offspring size as they relate to environmental conditions, adaptation, physiological constraints and evolutionary history (e.g., Bagenal, 1971; Thresher, 1982; Bernardo, 1996; Chambers, 1997). Large comparative datasets that are needed to detect general patterns are rare among tropical species of the type we consider here.

On a global scale, interspecific comparisons of egg-size in marine fishes and invertebrates often show latitudinal patterns (Bagenal, 1971; Collin, 2003; Moran and McAlister, 2009). Fish species from low latitudes mainly have small pelagic eggs that produce smaller hatchlings while most of those from higher latitudes have large demersal eggs and large hatchlings (Leis et al., 2013). Larger eggs and faster larval growth at higher latitudes may be related to shorter growing seasons (e.g., see Brown et al., 1998). A similar pattern is evident in marine invertebrates where species

with small eggs and planktotrophic larval development are more common at low latitudes and species with large eggs and non-feeding or benthic development are more common at high latitudes (Thorson, 1950; Collin, 2003). On a regional scale, egg size in species with planktonic feeding larvae has long been thought to vary with availability of planktonic food to support larval growth (e.g., Lessios, 1990). Under low productivity conditions females are thought to produce bigger eggs that in turn produce larger, better-provisioned, hatchlings that are better equipped to survive when there is less larval food available (Cushing, 1967; Bagenal, 1971). Data from neotropical invertebrates support this pattern. In Panama, geminate pairs of invertebrate species, which were formed when the rising isthmus of Panama split a single species (Jordan, 1908), display differences in egg size that have been attributed to differences in larval food supplies in the two oceans. The Caribbean has lower productivity than the tropical East Pacific, and Caribbean echinoderms, bivalve molluscs and bryozoans have volumetrically larger eggs or offspring than do their eastern Pacific congeners, a pattern consistent with the larval-food hypothesis (Lessios, 1990; Jackson and Herrera, 1999; Marko and Moran, 2002; Moran, 2004). However, the two coasts of Panama differ in a number of other environmental factors (e.g., salinity: **Table 1**; and see Alory et al., 2012), allowing for the

Table 1 | Variation in oceanographic conditions on the Caribbean and Pacific coasts of Panama where we collected fish eggs for this study.

Variable [Mean value]	Caribbean		Pacific	
	Cool season	Warm season	Cool season	Warm season
Temperature (°C)	27.9	28.7	25.5	28.1
(Range)	(27.1–30.0)	(26.6–30.0)	(18.7*–25.5)	(24.9–30.2)
Salinity (ppt)	34.8	34.2	32.2	29.2
Nitrate/Nitrite (μm)	0.23	0.29	1.03	0.27
Phosphate (μm)	0.04	0.04	0.37	0.12
Chlorophyll A (mg m^{-3})	0.36	0.41	1.48	0.59
Phytoplankton (cells ml^{-1})	60	100	663	166
Zooplankton (10^3 sample^{-1})	0.8	1.2	8.8	7.0

After Table 1 of D'Croz and Robertson (1997); note that warm and cool seasons in that paper differ slightly from those here; *falls as low as 14°C in some years.

possibility that other biotic or abiotic factors may have played a role in generating this pattern.

Wellington and Robertson (2001) compared egg-size and larval development among closely related pelagic spawning reef fishes on both sides of the isthmus of Panama, and transisthmian patterns of larval development (but not egg-size variation) among demersal spawners. They found that larval growth rates and pelagic larval duration were similar among closely related transisthmian species of both pelagic and benthic spawners. Because larval development is largely dependent on food availability and/or temperature (Houde, 1989; Heath, 1992; Houde and Zastrow, 1993; Chambers, 1997; Rilling and Houde, 1999; Bailey and Heath, 2001; Wilson and Meekan, 2002; Meekan et al., 2003), these transisthmian similarities raise questions about the extent to which food availability is the main determinant of transisthmian variation in egg size (Wellington and Robertson, 2001). In the present study we extend that work by assessing transisthmian and seasonal patterns of variation in egg-size among geminate and other congeneric demersal spawners, and include a broader range of species than were considered in Wellington and Robertson's (2001) study of larval development.

Intraspecific environmental variation in egg size in marine invertebrates and fishes is often linked to both temperature and productivity. The existence of such intraspecific plasticity is particularly useful in explorations of the adaptive significance of egg size because plastic responses to environmental variation are often considered to be adaptive (Padilla and Adolph, 1996; Hughes et al., 2003; see Einum and Fleming, 2002; for fish eggs in particular). In addition, unlike the situation for interspecific comparisons, intraspecific comparisons are not confounded by independent evolutionary histories. Temperate and tropical fishes in marine and freshwater environments commonly exhibit intraspecific latitudinal and seasonal variation in egg size similar to the interspecific patterns (Cushing, 1967; Bagenal, 1971; Marteinsdottir and Able, 1988; Clarke, 1989; Fleming and Gross, 1990; Chambers, 1997; Johnston and Leggett, 2002; Leis and

McCormick, 2002; Kokita, 2003). In fishes that produce pelagic larvae, egg size usually decreases with increasing primary production (a proxy for larval food supply) and with increasing temperature (Cushing, 1967; Bagenal, 1971; Chambers, 1997). In fact, egg size and offspring size vary inversely with temperature across a diverse array of metazoan taxa (Atkinson et al., 2001, e.g., see Fischer et al., 2003), a pattern referred to as the "offspring temperature-size rule (OTSR)."
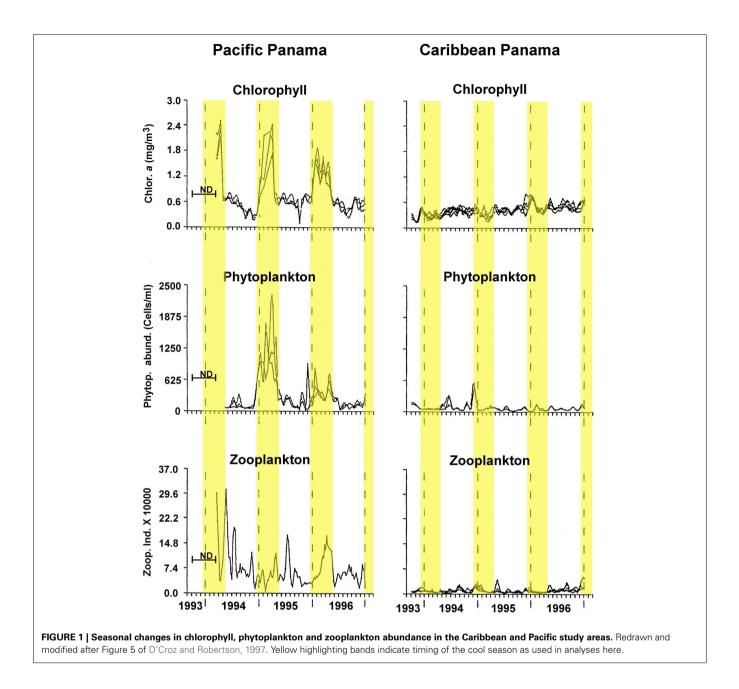
Here, we test the productivity hypothesis among neotropical reef fishes that have pelagic larvae by comparing not only egg-size among closely related species separated by the Isthmus of Panama, but also seasonal intraspecific variation in egg-size on both sides of the Isthmus. This extends a study of transisthmian patterns of larval growth and development among many of the same species (Wellington and Robertson, 2001). For the present analysis the most important differences in the environments of the two coasts of the Isthmus are as follows (**Table 1**; **Figure 1**): (i) The Pacific side of the Isthmus has a higher average level of primary production, a lower average temperature, and greater seasonal and short-term variation in both factors than does the Caribbean side. (ii) While there is a large peak of primary production, during the markedly cooler upwelling season on the Pacific coast, there is no clear seasonal change in primary production on the Atlantic coast, with only sporadic small increases intermittently throughout the year (D'Croz and Robertson, 1997; and see **Figure 1**, here.). This decoupling of seasonal temperature and productivity regimes presents an opportunity to disentangle effects of those two important factors.

In this paper examine the evolution of intra- and inter-specific variation in egg size of 24 species of tropical marine fishes from four different families, taking advantage of the varying environmental conditions and seasonal effects generated by the Isthmus of Panama. Based on hypothesized relationships between the various environmental factors and egg-size described above we made the following predictions about variation in egg size. If larval-food availability has an overriding influence on interspecific variation in egg size then (i) eggs of Pacific species should be smaller than those of Atlantic geminates and, perhaps, other congeners. Further, if larval food availability has an overriding influence on intraspecific variation in egg size then (ii) egg-size should be minimal during the cool season in Pacific species, and should show little or no seasonal change in Caribbean species. Alternatively, if temperature is of overriding importance, then (iii) Caribbean species should have smaller eggs than Pacific species, at least during the cool season, and (iv) within species, egg size should be inversely related to temperature on both sides of the Isthmus. It is possible that both environmental factors are involved in determining egg size, influencing patterns of variation in different ways and at different levels.

MATERIALS AND METHODS
SEA-TEMPERATURE DATA
Sea-temperature data were collected at 6 m depth in Pacheca Island (8.65°N, 79.05°W), in the Bay of Panama, and at 7 m depth at Punta de San Blas (9.45°N, 78.99°W), (**Figure 2**), on the Caribbean coast, by the Smithsonian Tropical Research Institute's Environmental Monitoring Program (see

FIGURE 1 | Seasonal changes in chlorophyll, phytoplankton and zooplankton abundance in the Caribbean and Pacific study areas. Redrawn and modified after Figure 5 of D'Croz and Robertson, 1997. Yellow highlighting bands indicate timing of the cool season as used in analyses here.

Kaufman and Thompson, 2005). Hourly or bi-hourly measurements were averaged for each week from 1992 to 2002 for the Pacific, and 1992 to 1999 for the Caribbean, and we used those weekly averages to define the two seasons (**Figure 3**). For the Caribbean, the cool season included the weeks when average temperature was below 28.5°C (weeks 1–15 and 51–53), with the remainder in the warm season. For the Pacific, cool season included the weeks when the average temperature was below 27°C (weeks 1–18 and 52–53).

STUDY SPECIES
We obtained egg-size data from 24 species representing seven genera and four families of neotropical reef fishes that produce benthic eggs and have pelagic larvae. These include three

pairs of transisthmian geminate species: one species pair of *Abudefduf* and one of *Chromis* (both Pomacentridae), and one of *Ophioblennius* (Blenniidae) (see Bermingham et al., 1997; Muss et al., 2001). The single Atlantic and Pacific representatives of *Acanthemblemaria* belonged to the same clade, but are not geminate sisters (Eytan et al., 2012). The single Pacific species of *Coryphopterus* is not the geminate sister of any of the three Caribbean species (J Van Tassell, pers. com., 2013). No phylogeny yet exists for *Microspathodon*, for which we sampled all three neotropical species. Finally, we sampled two of eight east Pacific species and six of seven Caribbean species of *Stegastes* (Pomacentridae). None of these constitute clearly identifiable geminate sister species pairs, although they belong to four genetic lineages (Rocha et al., unpublished data): lineages I and II include
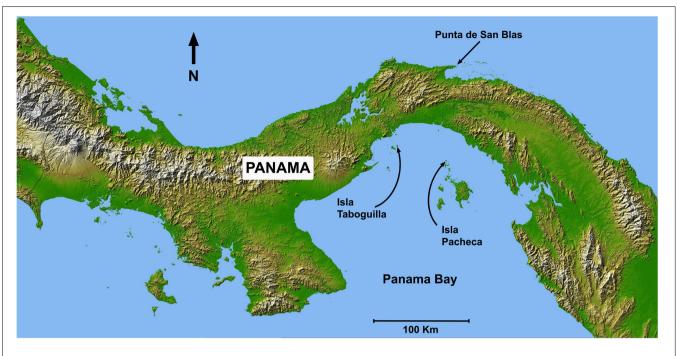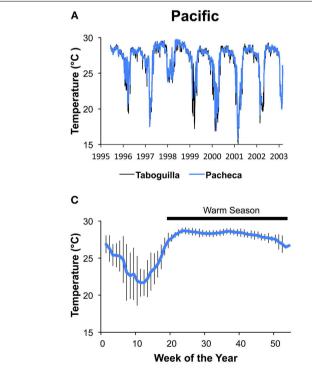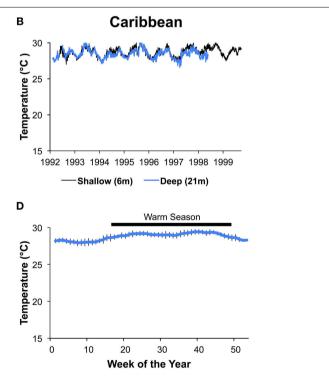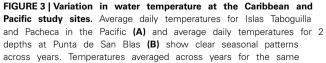
FIGURE 2 | Map of Panama showing Caribbean and Pacific study locations: Taboguilla Island (8.8°N, 79.51°W), Pacheca Island (8.65°N, 79.05°W), and Punta de San Blas (9.45°N, 78.99°W).

FIGURE 3 | Variation in water temperature at the Caribbean and Pacific study sites. Average daily temperatures for Islas Taboguilla and Pacheca in the Pacific **(A)** and average daily temperatures for 2 depths at Punta de San Blas **(B)** show clear seasonal patterns across years. Temperatures averaged across years for the same week show a colder and more variable temperatures during upwelling the Pacific depicted with data from Pacheca island **(C)** compared to the more stable Caribbean depicted with data from the shallow site at Punta de San Blas **(D)**. Vertical lines show standard deviations of the mean.

S. leucostictus and *S partitus* respectively, both Caribbean species whose clades lack a Pacific member; lineage III includes *S. planifrons* (Caribbean) and *S. acapulcoensis* (Pacific); and lineage IV includes *S. flavilatus* (Pacific), and *S adustus*, *S diencaeus* and *S. variabilis* (all Caribbean).

MODES OF REPRODUCTION OF THE STUDY SPECIES

All the Caribbean species in this study spawn and recruit pelagic larvae throughout the year, although spawning diminishes in many species during the cool season (Robertson, 1990). In the Bay of Panama, the study species spawn throughout most of the year, with a hiatus during the most intense periods of upwelling. Females of all species lay eggs in a close-packed monolayer on a hard nest surface, then leave, and the male cares for the eggs until hatching. Clutches laid on different days occupy different parts of the nest. In some *Stegastes* species multiple females contribute to a single clutch (e.g., Petersen, 1990) while in others only one female typically does so (e.g., *S. leucostictus*, Clifton and Robertson, unpublished data). In *Abudefduf* and *Chromis* species, each clutch usually is the product of spawning by multiple females, with several often spawning simultaneously in the same nest (Robertson, unpublished observations).

EGG COLLECTION

In most cases eggs were sampled during lunar peaks of spawning activity (see Robertson, 1990; Robertson et al., 1990), in both warm and cool seasons from 1983 to 1993. However, in four Caribbean species (*A. saxatilis*, *M. chrysurus*, *S diencaeus*, and *S. partitus*) ∼10 clutches per month were collected over a 3-year period. Eggs were collected at depths of 1-8 m, at Punta de San Blas (Caribbean coast), and at Taboguilla and Pacheca Islands (Pacific coast; see **Figure 2**).

Only clutches collected within several hours of being spawned (determined by inspection of newly collected clutches under a dissecting microscope) were used for egg-size assessments. Each sample comprised newly laid eggs collected from a single nest and, in most species, could therefore include eggs from multiple females. To facilitate collection of clean eggs, males of many species were provided with artificial nest substrata: clay tiles, sections of rigid fiberglass sheet, glass jars, and half-sections of PVC tubing. Eggs change color during development and fresh clutches are easy to distinguish from older clutches. Eggs were scraped off the nest surface underwater, sucked into a 20 ml syringe and kept on ice until size measurements were made. Maximum length and maximum width were measured for 20 undamaged eggs in each sample, using a calibrated graduated eyepiece in a Wild M5 dissecting microscope. The sample was then stored frozen until processed for weight and energy content.

In *Chromis multilineata* naturally spawned eggs invariably had large amounts of sand stuck to them, which precluded accurate measurements of egg-volume in this species. To obtain samples of clean eggs of this species for determination of egg content we speared females in the act of egg-laying, and gently stripped ripe eggs from them into vials within seconds of capture.

ESTIMATION OF EGG VOLUME, WEIGHT, AND ENERGY CONTENT

Most analyses of egg-size variation in marine fishes have been based on egg diameters or volumes, although egg dry weight and the size of newly hatched larvae have been used in some studies (Chambers, 1997; Kerrigan, 1997; Kokita, 2003; McCormick, 2003). Analyses of transisthmian variation in egg size among invertebrates have been based on either direct measurements of egg diameter or volume (Lessios, 1990; Marko and Moran, 2002), or indirect measurements of offspring size (Jackson and Herrera, 1999; Fortunato, 2004). Although egg volume may accurately predict differences in egg content among species of invertebrates (Jaeckle, 1995; Moran and McAlister, 2009) it does not necessary do so within species (McEdwards and Carson, 1987; McEdwards and Coulter, 1987). In most fishes, the embryo is surrounded in the egg by a large, water-filled perivitelline space (Craik and Harvey, 1984; Riss-Vestergaard, 2002). That space is produced by hydration of the egg, which occurs from shortly before spawning to shortly after fertilization (Tyler and Sumpter, 1996). Variation in the extent of hydration may weaken the relationship between egg volume and content. As all of our study species produce eggs with large perivitelline spaces, variation in egg volume may reflect different levels of hydration rather than maternal investment. Hence we examined egg volume, dry weight and energy content.

DETERMINATION OF AVERAGE EGG VOLUME, DRY WEIGHT, AND ENERGY CONTENT

We estimated the volumes of the non-spherical (oblate spheroid, ovoid or tapering cylindroid) eggs produced by the study species as follows. Camera lucida drawings were made of the outlines of five eggs from each of two clutches for each species. The sum of the volumes of 150 cylindrical subsections of each of those eggs was used to estimate its total volume. The average of those volumes expressed as the percentage of the volume of a cylinder with the same length and diameter was then calculated. This correction factor was used to estimate the average volume of an egg in each clutch of a species from measurements of maximum length and diameter of 20 eggs per clutch.

For determination of average dry weight and energy content of an egg in a clutch each sample of eggs was first carefully cleaned of all debris under a microscope, then rinsed ∼6 times in distilled water, and finally stored in a −80°C freezer. Subsequently each sample was lyophilized for at least 8 h and stored in a desiccator in a −80°C. To determine the average dry weight of an egg a subsample of ∼20 counted, intact dried eggs was weighed on a (daily calibrated) Cahn Model 28 Microbalance, after the desiccator containing the samples had been allowed to warm to room temperature in an air-conditioned room. After this the energy density (Joules mg^{-1} dry weight) of the sample was determined using a Phillipson Microbomb Calorimeter (Gentry Instruments) calibrated with a sample of Benzoic acid at the beginning of each day's measurements. Average energy content of an egg of each clutch was calculated as (average energy density of the clutch) × (the average dry weight of an egg in that clutch).

STATISTICAL ANALYSES

The correlation between the average egg volume, dry weight and energy content for each clutch was assessed using Pearson correlations, calculated with JMP®, Version 9.0.2. SAS Institute Inc., Cary, NC, 1989–2007, using the "Multivariate" menu. Due to

missing values Restricted Maximum Likelihood (REML) was used to calculate the significance of the correlations. For interspecific correlations between different measures of egg size, averages were calculated for warm season egg sizes for each species. Warm season data were used because the temperature conditions experienced then are more typical of the conditions over most of the year, and because the uneven sampling and small cool season sample sizes would unequally effect the species means if they were included. The intraspecific relationship between egg size and temperature was assessed in a similar way. We calculated the correlation between the average egg volume, weight or energy for each clutch with the average temperature recorded for that week.

Incomplete taxon sampling for diverse lineages, and low numbers of species in the fully sampled lineages precluded the use of phylogenetic comparative methods at either the genus– or global levels. Instead we use Two-Way ANOVA's to test for the effects of season and species, and the interaction between season and species on egg size for each genus. Most comparisons included a pair of species, one from the Pacific and one from the Caribbean, although a few involved more species. For *Stegastes*, the transisthmian comparisons were broken into two clades that each contained one Pacific species: (i) *S. acalpulcoensis* (Pacific) and *planifrons* (Caribbean), and (ii) *S. flavilatus* (Pacific) with, *S. adustus*, *S. diencaeus* and *S. variabilis* (Caribbean). Egg data were log transformed to reduce inequality among variances before ANOVA tests. *Post-hoc* Tukey's HSD pairwise comparisons were used to test for significant patterns when significant interactions were present, and power analysis was used to estimate the power of the test and to solve for the least significant number (i.e., the sample size needed for 50% power, given the raw effect size and the standard error of the residual error in the model) and the least significant value (i.e. the size of an effect that could produce a *p*-value of 0.05 given the sample size and the standard error of the residual error in the model). Because data on egg volume were available for many more dates than the other measures, analyses of egg volume were also conducted on a dataset of clutches for which egg weight data were also available. This was aimed at demonstrating whether any differences between results for egg volume and egg content are due to different sample sizes and/or different distributions of sampling dates.

RESULTS

RELATIONSHIPS BETWEEN EGG VOLUME AND CONTENT

Average egg volume, dry weight and energy content varied measurably among clutches from each species. Summary statistics of seasonal means of egg size variation for each species are presented in Supplementary Table 1. Overall, egg volume was not highly correlated with investment per egg within species. The Pearson correlation between average volume of an egg in a clutch and the average dry weight of an egg in the same clutch was statistically significant in only 14 of the 24 species, and volume was significantly correlated with average energy content of eggs for 12 of those 14 species (**Table 2**). The significant relationships were all positive but the correlation coefficient (*r*) for the relationship between egg volume and egg weight only exceeded 0.6 for one species, *Microspathodon dorsalis,* Egg volume was slightly more

closely correlated to egg energy content, with $r > 0.6$ for four species: *M. dorsalis, Chromis atrilobata, Stegastes partitus,* and *Ophioblennius macclurei.* In contrast to egg volume, egg weight was well correlated with egg energy content. The correlations were statistically significant in all species and *r*-values were typically high, with only five species showing an $r < 0.9$. The linear relationship between egg volume and the two measures of egg content are given in Supplementary Table 2.

Among species, the average species egg volume was an accurate predictor of both average species egg weight and average egg energy content (**Figure 4**). Linear regression of species averages from warm season measurements (the season for which the most data were available) show that among species egg volume was a good predictor of egg weight ($r^2 = 0.97$; $n = 24$; Egg Dry Weight $= 0.210$(volume) -0.001) and of egg energy content ($r^2 = 0.975$; $n = 22$; Egg Energy $= 5.04$ (volume) -0.04). Species from all but one genus, *Coryphopterus*, fall along the interspecific regression lines for egg volume vs. egg weight and egg energy content (**Figure 4**). *Coryphopterus* species fall on the same line as the other species when egg energy content is regressed against egg dry weight (**Figures 4E,F**).

PATTERNS IN EGG SIZE VARIATION BETWEEN SEASONS AND OCEANS

In general a Two-Way ANOVA analysis of egg size in each genus showed a pattern of larger egg size in Caribbean species compared to their Pacific relatives, and larger egg size in the cool season compared to the warm season in both oceans (**Figure 5**; **Table 3**; Supplementary Table 3). However, both patterns were not present in all genera for all of the measurements of egg size. When those patterns differed it was generally due to failure to find a significant effect in one or the other of the datasets and not due to different measures of egg size producing conflicting strongly supported results. This suggests that despite the poor correlations between egg volume and egg weight or energy, the overall patterns of variation in egg size can be recovered with any of these three measures of egg size.

All three measure of egg size were larger for the Caribbean species than for the Pacific species in *Acanthemblemaria*, *Coryphopterus*, *Ophioblennius*, and *Abudefduf* (**Table 3**; Supplementary Table 3; **Figures 5A,B,D,E**; on which significant Tukey's HSD values are indicated). In *Microspathodon* egg volume was larger in the Pacific species than in the Caribbean species and the other two measures did not differ significantly between the species (**Table 3**; **Figure 5G**). In *Stegastes*, the *S. acalpulcoensis-planifrons* pair followed the general rule that the Caribbean species produces larger eggs than the Pacific species (**Figure 5C**). In the *S. variabilis* clade eggs from the Pacific species (*S. flavilatus*) were smaller than those from each of the three Caribbean species (**Table 3**; **Figure 5H**). The two Caribbean species from the two clades that lack Pacific members, *S leucostictus* and *S. partitus* have the largest and smallest eggs, respectively, among the neotropical members of the genus (Supplemental Table 1). In *Chromis* egg dry weight and egg energy content were significantly lower in the Pacific species than in the Caribbean geminate (**Table 3**; **Figure 5**). In no case was there a clear-cut exception to the rule that Caribbean species have larger eggs in terms of content.

Table 2 | Pearson Correlations between average egg volume, egg dry weight, and egg energy content within species of the 24 neotropical reef fishes studied.
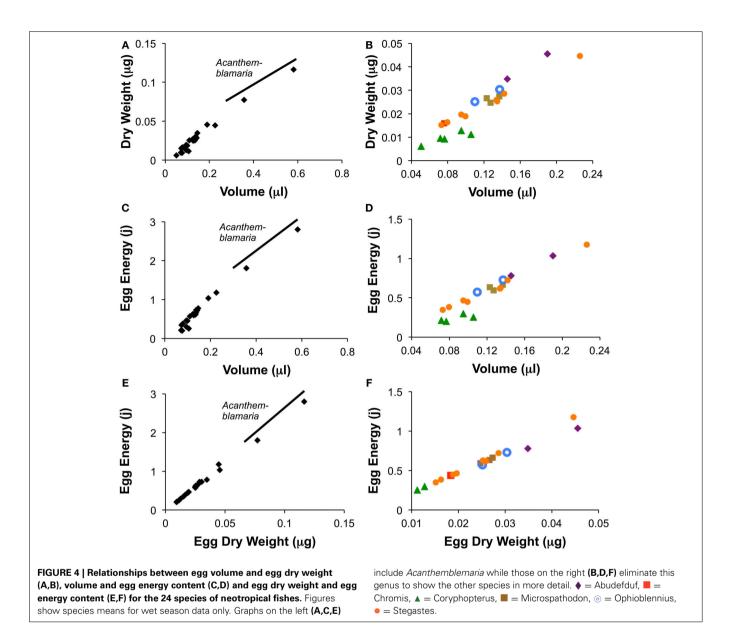
Species	Ocean	Egg volume vs. egg dry weight *r* (*n*)	Egg volume vs. egg energy *r* (*n*)	Egg dry weight vs. egg energy *r* (*n*)
POMACENTRIDAE				
Abudefduf saxatilis	Caribbean	−0.09 ns (66)	0.25 (48)	**0.43****(48)
Abudefduf troschelii	Pacific	**0.46***** (54)	**0.49***** (44)	**0.92*****(44)
Chromis multilineata	Caribbean	–	–	**0.56***(14)
Chromis atrilobata	Pacific	0.37 ns (15)	**0.90*** (6)	**0.92****(6)
Microspathadon bairdii	Pacific	0.20 ns (16)	−0.01 ns (10)	**0.95*******(10)**
Microspathodon chrysurus	Caribbean	0.25 ns (52)	**0.39*** (38)	**0.91*****(38)
Microspathodon dorsalis	Pacific	**0.73***** (32)	**0.75***** (24)	**0.98*****(24)
Stegastes adustus	Caribbean	**0.40**** (49)	**0.54**** (40)	**0.97*****(40)
Stegastes diencaeus	Caribbean	**0.55**** (30)	−0.02 ns (18)	**0.95*****(18)
Stegastes leucostictus	Caribbean	**0.42**** (56)	**0.48**** (41)	**0.58*****(41)
Stegastes partitus	Caribbean	**0.47***** (51)	**0.67***** (40)	**0.95*****(40)
Stegastes planifrons	Caribbean	**0.53*** (35)	0.44 ns (19)	**0.93*****(19)
Stegastes variabilis	Caribbean	**0.58***** (40)	**0.52****(24)	**0.93*****(24)
Stegastes. acapulcoensis	Pacific	0.23 ns (37)	0.32 ns (22)	**0.97*****(22)
Stegastes flavilatus	Pacific	**0.54***** (38)	**0.51**** (27)	**0.98*****(27)
BLENNIIDAE				
Ophioblennius macclurei	Caribbean	**0.47**** (43)	**0.89*****(28)	**0.90*****(28)
Ophioblennius steindachneri	Pacific	0.35 ns (24)	0.34 ns (24)	**0.97** ***(24)
CHAENOPSIDAE				
Acanthemblemaria rivasi	Caribbean	**0.56***** (39)	**0.47****(36)	**0.97*****(36)
Acanthemblemaria hancocki	Pacific	0.23 ns (46)	0.16 ns (42)	**0.83*****(42)
GOBIIDAE				
Coryphopterus dicrus	Caribbean	−0.40 ns (11)	−0.01 ns (11)	**0.82****(11)
Coryphopterus glaucofrenum	Caribbean	**0.47***** (48)	**0.54***(20)	**0.99*****(20)
Coryphopterus personatus	Caribbean	**0.38*** (28)	**0.41***(28)	**0.98*****(28)
Coryphopterus tortugae	Caribbean	−0.9 ns (27)	−0.13 ns (27)	**0.92*******(27)**
Coryphopterus urospilus	Pacific	**0.53***** (45)	**0.25**** (30)	**0.90*****(30)

Probability values (p): ns> 0.05;* < 0.05;** < 0.01;*** < 0.001. Calculated with JMP using the "Multivariate" menu. Due to missing values REML was used to calculate correlations. ND, no data.

Values in bold are significant at p < 0.05−0.01.

Eggs from the cool season had statistically significantly larger volumes, dry weight, and energy content than those collected in the warm season for all genera except for: *Chromis* and *Acanthemblemaria* (**Figures 5B,F**; **Table 3**). For the two *Chromis* species there was no significant effect of season on dry weight. Power analysis indicates that a sample size of over 300 would be necessary to detect significance of the very small size indicated by our data (Supplemental Table 3). For *C. atrilobata*, the single species for which we have egg volumes, there was likewise no effect of season on egg volume. Adequate samples were not available to test for a seasonal effect on egg energy content for *Chromis*. For *Acanthemblemaria*, both egg dry weight and egg energy content were larger during the cool season, but there was no significant effect of season on egg volume. No species showed any tendency, statistically significant or otherwise, toward having smaller eggs (volume or content) in the cool season.

In five of seven genera there was a significant interaction between season and species (**Figure 5**; **Table 3**). In all cases when

an interaction was present it showed a larger impact of season on egg size in the Pacific than in the Caribbean. For *Abudefduf* there was a significant effect of season on both egg weight and egg energy in the Pacific species, but not the Caribbean species (**Table 3**; Supplemental Table 3). In *Microspathodon* all species showed a significant effect of season on egg volume (a larger effect in the Pacific species; **Figure 5G**), but a significant effect on egg weight and energy content only in the Pacific species. For both clades of *Stegastes* egg volume showed a significantly stronger seasonal effect in the Pacific than the Caribbean (**Figures 5C,H**), but there was no significant interaction when egg weight or energy are analyzed. *Ophioblennius* showed a significant effect of season on egg volume in the Pacific species only (**Figure 5E**), but no interaction effect on egg weight or energy content (**Table 3**; Supplementary Table 3). Nor did that effect on volume remain in the analyses of clutches for which data on both volume and content were available. This suggests that the lack of a statistically significant interaction term

FIGURE 4 | Relationships between egg volume and egg dry weight (A,B), volume and egg energy content (C,D) and egg dry weight and egg energy content (E,F) for the 24 species of neotropical fishes. Figures show species means for wet season data only. Graphs on the left **(A,C,E)** include *Acanthemblemaria* while those on the right **(B,D,F)** eliminate this genus to show the other species in more detail. ♦ = Abudefduf, ■ = Chromis, ▲ = Coryphopterus, ■ = Microspathodon, ⊙ = Ophioblennius, ● = Stegastes.

in the analysis of egg weight or energy in this case is due to either small sample sizes or the distribution of dates sampled. The *Acanthemblamaria* species showed no significant difference in their response to season (**Table 3**; Supplemental Table 3; **Figure 5B**) and the *Coryphopterus* species also showed a very similar response of egg volume to season, although there was a marginally significant interaction between season and species (**Table 3**; **Figure 5D**).

EGG-SIZE vs. TEMPERATURE

There was a general tendency toward a decline in egg size with temperature across the entire set of species (**Table 4**). There were negative correlations between the average temperature for the week and egg volume in 18 of 24 species. Fifteen of the species showed a negative correlation between temperature and egg dry weight and 13 showed a negative correlation between temperature and egg energy content. Only four species did not

show a significant correlation in any of the three measures: *C. atralobata* and *M. bairdii* from the Pacific and *C. tortugae* and *C. dicrus* from the Caribbean. The only species to show a significant positive correlation between egg energy content and temperature was the Pacific *S. acapulcoensis*. Measurements covered only samples from the wet season, and correlations for this species therefore are not comparable to the correlations for other Pacific species that covered the much larger temperature range over both seasons. Overall, the statistically insignificant relationships between temperature and egg volume or content, and non-significant relationships between temperature and egg dry weight or egg volume were typically associated with small sample sizes and/or reduced temperature ranges across which eggs were sampled. For example, power analysis suggests that for *C. atrilobata* a sample size of 48 (compared to our 21) could be sufficient to detect a significant effect of temperature on egg volume).

FIGURE 5 | Results of the Two-Way ANOVA showing the difference between species, the effect of season, and the interaction between the two facts on egg volume (μl). Results show the interaction plots from the 19 species included in Two-Way ANOVA. The Caribbean species is indicated in blue and the Pacific in red. Letters next to the data points indicated the results of the *post-hoc* Tukey's HSD test, with values not connected by the same letter differing significantly from each other. Error bars indicate the standard deviation of the log-transformed data on which the analyses were conducted. The un-transformed means are presented for ease of comparison with other studies and the tables. **(A)** *Abudefduf*, **(B)**. *Acanthemblamaria*, **(C)** *Stegastes (clade iii)*, **(D)** *Coryphopterus*, **(E)** *Ophioblennius*, **(F)** *Chromis*, **(G)** *Microspathadon*, and **(H)** *Stegastes (clade iv)*.

DISCUSSION

RELATIONSHIPS BETWEEN EGG VOLUME, WEIGHT, AND ENERGY CONTENT

We found that the relationship between dry weight and egg energy content is almost perfectly linear across the 24 tropical reef species studied (**Figure 4**). This suggests that these eggs are broadly similar in composition of lipids, proteins and carbohydrates. Egg volume is also a good predictor of egg dry weight and egg energy content among species, but these relationships are not quite so uniform across the genera. Across the entire range of volumes the relationship is fairly good. However, examining egg volumes under 0.25 μl shows that the five species of *Coryphopterus* have a more gradual increase in dry weight and energy content with volume than do the other genera. Since *Coryphopterus* species are not outliers in the correlation between dry weight and energy (**Figures 4E,F**), this suggests that species in this genus have more

highly hydrated eggs than eggs from the other groups. This may relate to natural nests of *Coryphopterus* species typically being in narrow-mouthed sandy burrows in the substratum in deeper habitat with no wave action. In such poorly ventilated situations relatively large egg surface-area may be important for embryo respiration.

Within species the relationship between egg volume, dry weight, and energy content is not so clear-cut. We found that, in general, the average volume of an egg is typically a poor predictor of the average weight or energy content of an egg in the same clutch (**Table 2**; Supplementary Table 2). Average dry weight of an egg in a clutch was however almost always a very good predictor of the average energy content of an egg in the same clutch. This applies irrespective of whether one or more females typically contribute to a clutch in the species examined. These relationships indicate that the extent of hydration (which effects volume but not dry weight or energy content) of an egg is not closely linked to the amount of material it contains. The close correlation between the average dry weight of an egg compared to the average energy content of an egg from the clutch also suggests that within clutch variation in egg content does not drive the poor relationship between egg volume and content. Volume/content relationships have been found to be stronger in interspecific comparisons than in within population comparisons in other fishes (Chambers, 1997), so this may be typical of fishes in general.

Despite the weakness of the volume/content relationship within species, seasonal differences in egg volume, dry weight, and energy content did follow the same general pattern in most of the species. When different results were obtained from analyses using these different proxies for egg size it was largely due to a failure to detect a significant effect in the datasets with small sample sizes. In no cases did the results from using egg volume differ significantly in direction from the results of the same analysis using egg dry weight. Thus, despite the large amount of unexplained variation in volume/content relationships within a single population, egg volume, which is significantly easier to measure than dry weight or energy content, does represent a useful eggsize indicator for comparing seasonal variation in egg-content as well as differences in egg-content among these closely related reef fishes.

TRANSISTHMIAN VARIATION IN EGG-SIZE

We hypothesized that if larval-food availability has an overriding influence on interspecific variation in egg size then eggs of Pacific species should be smaller than those of Atlantic geminates and, perhaps, other congeners. If temperature is of overriding importance, then Caribbean species should have smaller eggs than Pacific species, at least during the cool season, when Pacific temperatures are much lower. Our data show that eggs of Pacific members of seven genera from four families of reef fishes typically are smaller in volume and content than those of geminate or other congeneric Caribbean species, consistent with the hypothesis that productivity and not temperature is driving the differences between the two coasts. These differences parallel transisthmian differences in the sizes of hatchling larvae of many of the same fish species (Wellington and Robertson,

Table 3 | Results of the Two-Way ANOVA of 3 different measurements of egg size.

Genus	Effect of season	Effect of species	Season × species	r^2 and N for the model
POMACENTRIDAE				
Abudefduf saxatilis vs. A. troschelii				
Log egg volume	Dry > wet***	Carib > Pac ***	ns	0.73, 457
Log egg dry weight	Dry > wet**	Carib > Pac ***	Pac: seasonal effect; Carib: no seasonal effect *	0.70, 120
Log egg energy	Dry > wet***	Carib > Pac ***	ns	0.67, 92
Culled log egg volume	Dry > wet***	Carib > Pac ***	Pac: seasonal effect; Carib: no seasonal effect *	0.86, 120
Chromis multilineata vs. C. atrilobata				
Log egg volume	ns (*C. atrilobata* only)	–	–	–
Log egg dry weight	Ns	Carib > Pac***	ns	0.48, 30
Log egg energy	–	Carib > Pac***	–	–
Microspathodon bairdii, and M. dorsalis vs. M. chrysurus[#]				
Log egg volume	Dry > wet***	Pac > Carib***	All: seasonal effect. Carib effect < Pac effect.***	0.48, 465
Log egg dry weight[&]	Dry > wet***	ns	Pac: seasonal effect; Carib: no seasonal effect. **	0.24, 84
Log egg energy[&]	Dry > wet***	ns	Pac: seasonal effect; Carib: no seasonal effect*	0.26, 62
Culled log egg volume	Dry > wet***	Pac > Carib***	All: seasonal effect. Carib effect < Pac effect.***	0.56, 115
Stegastes variabilis, S. adustus and S. diencaeus vs. S. flavilatus				
Log egg volume	Dry > wet***	Carib > Pac ***	All: seasonal effect. Carib effect < Pac effect. ***	0.88, 620
Log egg dry weight	Dry > wet*	Carib > Pac ***	ns	0.57, 157
Log egg energy	Dry > wet*	Carib > Pac ***	ns	0.63, 109
Stegastes acalpulcoensis vs. S. planifrons				
Log egg volume	Dry > wet***	Carib > Pac ***	All: seasonal effect. Carib effect < Pac effect. ***	0.81, 135
Log egg dry weight	Dry > wet*[$]	Carib > Pac ***	ns	0.60, 73
BLENNIIDAE				
Ophioblennius macclurei vs. steindachneri				
Log egg volume	Dry > wet***	Carib > Pac ***	Pac: seasonal effect; Carib: no seasonal effect***	0.65, 137
Log egg dry weight	Dry > wet***	Carib > Pac ***	ns	0.59, 67
Log egg energy	Dry > wet***	Carib > Pac ***	ns	0.73, 52
Culled log egg volume	Dry > wet***	Carib > Pac ***	ns	0.78, 67
CHAENOPSIDAE				
Acanthemblemaria rivasi vs. hancocki				
log egg volume	ns	Carib > Pac ***	ns	0.92, 90
log egg dry weight	Dry > wet***	Carib > Pac ***	ns	0.81, 82
Log egg energy	Dry > wet***	Carib > Pac ***	ns	0.84, 78
GOBIIDAE				
Coryphopterus glaucofrenum vs. C. urospilus				
Log egg volume	Dry > wet***	Carib > Pac ***§	All: seasonal effect. Carib effect < Pac effect.*	0.28, 117
Log egg dry weight	Dry > wet***	Carib > Pac ***§	ns	0.50, 92
Log egg energy	–	–	–	

*Comparisons between species and seasons for egg volume, egg weight, and egg energy content for geminate species or close relatives in each genus. Egg data were log transformed to reduce inequality among variances. *p < 0.05; **p < 0.01; ***p < 0.001; ns, p > 0.05. In the cases where the dataset for egg volume includes many more dates and much larger sample sizes than weights or energy content, the volume datasets were also "culled" to include only the dates for which the egg weight and egg energy content were also available.*

§One-Way ANOVA on all species for the wet season only shows the Pacific species is significantly smaller than all of the Caribbean species.

#Two outliers in the volume measurements removed.

$Becomes significant at p < 0.05 when the non-significant interaction is removed.

&Does not include M. bairdii.

2001) as well as transisthmian differences in egg-volume among echinoderms (Lessios, 1990) and bivalve molluscs (Marko and Moran, 2002; Moran, 2004) and proxies for hatching size in bryozoans (Jackson and Herrera, 1999) and molluscs (Fortunato, 2004).

While our data and similar transisthmian data on marine invertebrates support the larval-food hypothesis other scenarios cannot yet be ruled out. For example in food-rich environments growth rates are increased and predation pressure may be reduced making the rapid attainment of a size refuge from

Table 4 | Pearson Correlations between average weekly temperature across years and egg volume, egg dry weight, and egg energy content within species of the 24 neotropical reef fishes studied.

Species	Ocean	Weekly temperature vs. egg volume r (n)	Weekly temperature vs. egg dry weight r (n)	Weekly temperature vs. egg energy r (n)
POMACENTRIDAE				
Abudefduf saxatilis	Caribbean	**−0.250 (374)*****	**−0.297 (66)***	**−0.348 (48)***
Abudefduf troschelii	Pacific	**−0.40 (83)*****	−0.245 (54) ns	**−0.363 (44)***
Chromis multilineata	Caribbean	–	−0.422 (15) ns	−0.068 (14) ns
Chromis atrilobata	Pacific	−0.293(21) ns	−0.223 (15) ns	−0.580 (6) ns
Microspathadon bairdii	Pacific	−0.274 (31) ns	0.248 (16) ns	0.340 (10) ns
Microspathodon chrysurus	Caribbean	**−0.472 (350)*****	**−0.302 (52)***	−0.245 (38) ns
Microspathodon dorsalis	Pacific	**−0.683 (86)*****	**−0.730 (32)*****	**−0.846 (24)*****
Stegastes adustus	Caribbean	**−0.618 (77)*****	**−0.347 (49)***	**−0.418 (40)****
Stegastes diencaeus	Caribbean	**−0.335 (338)*****	**−0.394 (30)***	0.382 (18) ns
Stegastes leucostictus	Caribbean	**−0.326 (91)****	**−0.589 (56)*****	−0.235 (41) ns
Stegastes partitus	Caribbean	**−0.377 (361)*****	**−0.469 (51)*****	**−0.593 (40)*****
Stegastes planifrons	Caribbean	**−0.686 (36)*****	**−0.383 (36)***	**−0.542 (19)***
Stegastes variabilis	Caribbean	**−0.418 (65)*****	**−0.595 (40)*****	**−0.483 (24)***
Stegastes. acapulcoensis	Pacific	**−0.850 (99)*****	−0.053 (37) ns	**0.498 (22)***
Stegastes flavilatus	Pacific	**−0.875(140)*****	**−0.563 (38)*****	**−0.521 (27)****
BLENNIIDAE				
Ophioblennius macclurei	Caribbean	**−0.423 (54)****	**−0.732 (43)*****	**−0.834 (28)*****
Ophioblennius steindachneri	Pacific	**−0.715 (83)*****	**−0.445 (24)***	**−0.471 (24)***
CHAENOPSIDAE				
Acanthemblemaria rivasi	Caribbean	−0.031 (40) ns	**−0.609 (39)*****	**−0.537 (36)*****
Acanthemblemaria hancocki	Pacific	0.238 (50) ns	**−0.530 (46)*****	**−0.447 (42)****
GOBIIDAE				
Coryphopterus dicrus	Caribbean	−0.058 (14) ns	0.125 (11) ns	−0.016 (11) ns
Coryphopterus glaucofrenum	Caribbean	**−0.441 (65)*****	**−0.472 (48)*****	−0.126 (20) ns
Coryphopterus personatus	Caribbean	**−0.425 (28)***	0.034 (28) ns	−0.047 (28) ns
Coryphopterus tortugae	Caribbean	0.084 (29) ns	0.121 (27) ns	0.104 (27) ns
Coryphopterus urospilus	Pacific	**−0.651 (53)*****	**−0.733 (45)*****	**−0.532 (30)****

Probability values (p): ns > 0.05; *<0.05; **<0.01; ***<0.001. Calculated with JMP using the "Multivariate" menu. Due to missing values REML was used to calculate correlations.

Values in bold are significant at p < 0.05−0.01.

predation less important. This may result in the production of smaller and more numerous offspring when primary productivity is high. One obvious abiotic difference between the oceans, salinity (see Alory et al., 2012), is unlikely to explain our observed pattern as low salinity generally correlates with larger egg size or offspring size (Solemdal, 1967; Vallin and Nissling, 2000; Gimenez and Anger, 2001; and see Chambers, 1997), the opposite of our observed inter-oceanic and seasonal differences. Other abiotic factors that also differ strongly in inshore waters on the two sides of the Isthmus of Panama and might also affect reproductive activity of fishes include natural ocean acidification (e.g., Manzello et al., 2008) and reduced O_2 levels (D'Croz and O'Dea, 2007) in surface waters associated with upwelling activity along the Pacific coast of Panama. Recent manipulative experiments have assessed effects of ocean acidification, sometimes in association with increased temperatures, on egg development and early life histories of marine fishes. Those manipulations resulted in little or no effect on egg size and development and hatchling-size of various tropical and temperate fishes (Munday et al., 2009; Franke and Clemmensen, 2011; Miller et al., 2012; Hurst et al., 2013), although acidified conditions can either increase (Munday et al., 2009) or decrease larval growth (Franke and Clemmensen, 2011). Acidification can also stimulate higher reproductive output, perhaps at cost to egg volume and hatchling size (Miller et al., 2013). What significance this work has to predict potential effects of upwelling on Tropical Eastern Pacific fishes is unclear, as none of those acidification experiments examined effects of reduced pH at lower temperatures and O_2 levels, a combination characteristic of upwelling conditions.

SEASONAL VARIATION IN EGG-SIZE

We hypothesized that if larval food availability has an overriding influence on intraspecific variation in egg size then egg-size should be minimal during the cool season in Pacific species, and show little seasonal pattern of change among Caribbean species. If, on the other hand, temperature has an over-riding effect then seasonal variation in egg size should be negatively related

to temperature on both sides of the Isthmus. Our data show that none of the Pacific species produced smaller eggs (volume and content) during the cooler, higher productivity season, and that egg size changed seasonally in Pacific species and in some Caribbean species despite the lack of distinct seasonal change in productivity in the Caribbean. Thus, the hypothesis that seasonal change in egg size is an adaptive response to variation in the supply of larval food (Cushing, 1967; Hempel and Blaxter, 1967; Bagenal, 1971) is not supported by our data, nor by data from Mediterranean (Daoulas and Economou, 1986) and Hawaiian (Clarke, 1989) fishes.

Species from both sides of the Isthmus show a significant decrease of egg size with increasing temperatures. The patterns of cool vs. warm seasonal change in egg size as well as the negative correlation between egg size and average weekly temperature support the idea that temperature has an overriding effect on egg size variation within species. The significant interactions between season and species in our Two-Way ANOVAs support the role of temperature in controlling intraspecific variation in egg size. Six out of nine comparisons between Pacific and Caribbean pairs showed a significant interaction between ocean (i.e., species) and season. In all of these the Pacific species showed either a larger effect of season than did the Caribbean species, or no effect of season was recovered for the Caribbean (**Table 3**). Thus, the Pacific, the site with the largest temperature change across the seasons, has the largest change in egg size across the seasons, while species from the Caribbean which has a minimal change in temperature show smaller or no changes in egg size with season. These patterns are consistent with evidence from various marine metazoan taxa that egg size and offspring size virtually always show a negative correlation with environmentally relevant temperatures (Atkinson et al., 2001). This includes Panamanian slipper limpets, which produce larger diameter eggs and larger hatchlings when reared at cooler temperatures (Collin, 2012).

POTENTIAL EFFECTS OF MATERNAL CONDITION AND SIZE ON SEASONAL VARIATION IN EGG SIZE?

Maternal condition and size are well known to influence variation in egg-size in fishes (Chambers and Leggett, 1996; Kerrigan, 1997; Heath and Blouw, 1998; Kokita, 2003; McCormick, 2003), with larger females or those in better condition producing larger eggs. Could the seasonal patterns of variation in egg size and content that we found be due to either maternal factor? On both sides of the isthmus spawning is reduced during the cool period, often severely so on the Pacific coast. This suggests that environmental conditions during the cool season on both coasts are stressful for reef fishes. On the Caribbean coast strong, persistent onshore trade winds produce turbid conditions, and stronger surge and current flows on shallow reefs. On the Pacific side, cool season upwelling produces very low temperatures, reduced oxygen levels and reduced pH (see **Table 1**; D'Croz and O'Dea, 2007; Manzello et al., 2008). In the Pacific spawning often ceases during intensive upwelling, but continues at a reduced level during El Niño events, when upwelling activity is weaker and temperatures are higher than normal (Robertson, 1990). It seems unlikely that environmental conditions that inhibit spawning would be associated with improved female condition, resulting in the observed pattern of

larger eggs during the cool season. It also seems unlikely that seasonal variation in female size produces such seasonal variation in egg size. Those of our study species for which demographic data are available live for at least several years (Meekan et al., 2001; Caldow and Wellington, 2003). In addition, larval recruitment of those species is spread throughout most of the year (Robertson, 1989, 1990, 1992). A combination of these two characteristics would provide a year-round mixture of sizes of breeding females. At this point there is little support for the idea that seasonal variation in maternal condition and size drive the seasonal pattern of variation in egg size, although this hypothesis remains to be tested explicitly.

TEMPERATURE EFFECTS ON EGG SIZE: A BYPRODUCT OF PHYSIOLOGICAL CONSTRAINTS OR ADAPTIVE VARIATION?

On a species by species basis negative relationships between temperature and offspring size have often been explained as adaptive and involving specific aspects of each species' natural history. However it seems unlikely that such a diversity of organisms, phylogenetic histories and ecologies would consistently produce the same negative relationship (cf. Atkinson et al., 2001). For example, our observations show a consistently negative relationship between temperature and egg size among species in different families and suborders of fishes, taxa with a range of adult and larval ecologies that live in two oceans with differing oceanographic conditions. Species in some of those taxa exhibit a variety of patterns of seasonal change in larval recruitment with no consistent relationship to environmental factors thought to be primary determinants of larval survivorship (Robertson, 1990).

The ubiquity of the egg size temperature relationship has prompted the proposal that physiological constraints underlie the OTSR. The rates of growth and development of eggs may be affected differently by temperature, and if the temperature coefficient (Q_{10}) of egg-development rate is greater than that of egg-growth rate, this difference could lead to larger eggs at colder temperatures (e.g., Zuo et al., 2012). This idea has been proposed to explain the negative relationship between temperature and egg size for fishes (Chambers, 1997; Daoulas and Economou, 1986) and butterflies (Steigenga and Fischer, 2007). A similar constraint hypothesis, that all cells are larger at cooler temperatures and that eggs are simply just another cell type (Van Voorhies, 1996), could be tested by measuring the seasonal patterns of variation in size of both eggs and other cell types with fast turn-over rates, or by manipulating temperature in a laboratory setting to control for effects of other environmental cues. Concurrent, concordant temperature related variation in the size of eggs and other cells would lend support to Van Voorhies' hypothesis. Such an approach has provided some evidence of increased size in somatic cells with decreasing temperatures in other ectotherms (Azevedo et al., 2002; Arendt, 2007). Temperature also has effects on hormone systems that influence reproduction (Fraser et al., 2002) as well as oogenesis and egg-size and hatchling size (McCormick, 1999). Other mechanisms for physiological constraints include allometric differences of oxygen diffusion and metabolic rate in embryos or egg masses (Woods, 1999; Woods and Hill, 2004).

The physiological constraint hypothesis implies that variation in egg size with temperature is non-adaptive, at least with respect

to effects on embryonic or larval survivorship. As Chambers (1997) pointed out, deviations from this pattern could be particularly informative, because they could indicate an adaptive response. In the species we examined there is a large amount of variation in egg size. In these tropical species, particularly the Caribbean species which live in a low seasonality environment, and in which there are no consistent connections between larval survivorship and environmental variation (Robertson, 1990; Wellington and Robertson, 2001), seasonal egg-size variation may have no adaptive significance. There is accumulating evidence that, in diverse taxa of fishes from very different environments, seasonal variation in egg size does not offset the effects of environmental change on larval survivorship (Daoulas and Economou, 1986; Clarke, 1989; and see Robertson, 1990). Although some species appear to closely fit idealized relationships between egg size, productivity and temperature, a broad-based interspecific comparative approach assessing as many aspects of reproduction and environmental parameters as possible is necessary to understand what aspects of intraspecific variation in egg size are adaptive and what are due to phylogenetic or physiological constraints (Leis et al., 2013).

Although it may be unlikely that a diversity of organisms, phylogenetic histories and ecologies would produce a consistent OTSR, adaptive hypotheses still need to be considered. Such hypotheses range from general explanations relevant to a broad range of unrelated taxa with differing modes of reproduction, to those specifically tailored to a particular mode of reproduction and environmental circumstance. As with any kind of phenotypic plasticity, the OTSR may have originated from a physiological constraint, and subsequently been modified by selection to become adaptive (Eshel and Matessi, 1998; Hughes et al., 2003).

A general hypothesis relates to size-dependent effects of temperature on larval performance and survival. Small larvae from small eggs experience reduced survivorship in some fishes (e.g., Iguchi and Yamaguchi, 1994). Small fish larvae have reduced swimming performance under colder conditions due to temperature effects on water viscosity, and the metabolic cost of swimming increases at low temperatures (Hunt von Herbing, 2002). These effects may be too small in warm waters to be of significance to fishes on the Caribbean coast of Panama, and other consistently warm areas. However, seasonal changes in temperature in eastern Pacific upwelling zones may be sufficiently great (10–15°C) to affect larval performance, and survivorship in a size-dependent manner among fishes and other marine organisms. Production of larger eggs at colder temperatures might compensate for this effect. The fact that the Pacific species with the largest eggs shows almost no seasonal change in egg size (**Figure 5B**) offers support for this view.

A more specific hypothesis is relevant to some of our study species on both sides of the Isthmus. Clarke (1989) found that the negative relationship between egg- and hatchling-size and temperature in a tropical anchovy is adaptively linked to temperature effects on development of the egg to hatching: Temperature controls the rate of development of the larva in the egg and, hence, when it becomes competent to begin feeding. As the transition to first-feeding is a critical stage in the life history of pelagic fish larvae, and the larvae of this anchovy feed only during the day, a

failure to reach the most appropriate stage of development at the stage of the diel cycle that maximizes first-feeding opportunities may increase mortality of new hatchlings. Hence a combination of seasonal variation in the time of day of spawning and egg-size may allow developing larvae to achieve competency as the window of opportunity for first feeding arrives, regardless of the season (Clarke, 1989).

This "egg-development window" hypothesis could apply to some of our study species in the following way. In *Stegastes* species, at least, hatching appears to be restricted to a short period just after sunset, while spawning in some of the same species for which data are available occurs in the early morning (Petersen, 1990; Petersen and Hess, 1990; Robertson et al., 1990). In these species eggs develop rapidly and hatch 3–5 days after spawning. A failure to achieve an appropriate state of development during the hatching window would result in the larva spending an additional day in the egg, a substantial delay that could increase mortality through loss of eggs in the nest and through consumption of larval energy reserves. On the Caribbean coast of Panama, where temperatures vary seasonally by only ∼3°C, clutches of some of our *Stegastes* species do sometimes remain in the nest a day longer in the cool season than in warm season (Robertson, unpublished observations). Hence the prospect of a 1-day delay in hatching is real, even in this low seasonality environment. Adaptive seasonal variation in egg-size could ensure that larvae are at an appropriate stage of development when a narrow hatching window arrives regardless of the season.

There is tremendous variation among fishes in reproductive characteristics: in average egg-size; whether eggs are demersal, brooded or pelagic; whether or not there is parental care; to what extent there are spawning/hatching/first-feeding windows of opportunity; and the duration and timing of spawning seasons. Various formulations of the developmental-windows hypothesis that reflect this diversity of reproductive characteristics have the potential to account for a range of seasonal and latitudinal patterns of variation in egg-size. This hypothesis is more likely to be applicable to tropical species whose eggs develop rapidly than temperate species that take much longer to complete development not only because precise adjustments to the developmental sequence likely will be less feasible in the latter, but also because the relative energetic costs of a delay likely will be lower.

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SUPPLEMENTARY MATERIAL

The Supplementary Material for this article can be found online at: http://www.frontiersin.org/journal/10.3389/fevo.2014.00084/abstract

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CORRIGENDUM. (February 20, 2019): The original version of this Table published on January 7, 2015 as a supplement to this paper incorrectly listed units of Egg Dry Weight as micrograms (µg) rather than milligrams (mg). The units are listed as mg in this corrected version of the Table, below.

Supplementary Table 1: Summary of egg volume, dry weight, and energy content of eggs of 24 neotropical reef fishes averaged for the cool and the warm seasons.

Species	Ocean	Season	Egg Volume (µl)			Egg Dry Weight (µg mg)			Egg Energy (Joules)		
			Mean	St Dev	N	Mean	St Dev	N	Mean	St Dev	N
POMACENTRIDAE											
Abudefduf saxatilis	Caribbean	Warm	0.190	0.012	259	0.045	0.0024	45	1.034	0.071	31
		Cool	0.199	0.013	115	0.046	0.0028	21	1.075	0.047	17
Abudefduf troschelii	Pacific	Warm	0.146	0.007	45	0.035	0.0035	32	0.780	0.092	22
		Cool	0.153	0.010	38	0.038	0.0044	22	0.861	0.105	22
Chromis multilineata	Caribbean	Warm	-	-	-	0.018	0.002	8	0.438	0.020	7
		Cool	-	-	-	0.019	0.001	7	0.443	0.051	7
Chromis atrilobata	Pacific	Warm	0.076	0.002	14	0.016	0.001	8	-	-	-
		Cool	0.078	0.006	7	0.016	0.001	7	0.346	0.043	6

Microspathadon bairdii	Pacific	Warm	0.137	0.015	27	0.027	0.0037	14	0.665	0.075	10
		Cool	0.157	0.004	4	0.027	0.0075	2	-	-	-
Microspathodon chrysurus	Caribbean	Warm	0.123	0.007	246	0.027	0.004	35	0.634	0.086	21
		Cool	0.132	0.006	104	0.028	0.003	17	0.674	0.091	17
Microspathodon dorsalis	Pacific	Warm	0.127	0.012	37	0.025	0.0035	20	0.594	0.046	16
		Cool	0.152*	0.018*	47*	0.031	0.0033	12	0.751	0.083	8
Stegastes adustus	Caribbean	Warm	0.095	0.008	55	0.020	0.0046	34	0.465	0.116	25
		Cool	0.105	0.008	22	0.021	0.0012	15	0.518	0.030	15
Stegastes diencaeus	Caribbean	Warm	0.142	0.010	261	0.029	0.0039	28	0.722	0.054	16
		Cool	0.150	0.010	77	0.026	0.0009	2	0.640	0.028	2
Stegastes leucostictus	Caribbean	Warm	0.226	0.014	57	0.045	0.006	37	1.175	0.118	22
		Cool	0.238	0.016	34	0.051	0.0028	19	1.227	0.096	19
Stegastes partitus	Caribbean	Warm	0.073	0.008	252	0.015	0.0027	28	0.349	0.083	18
		Cool	0.079	0.005	109	0.017	0.0012	23	0.401	0.032	22

Stegastes planifrons	Caribbean	Warm	0.133	0.007	23	0.026	0.0032	22	0.617	0.071	10
		Cool	0.147	0.006	14	0.029	0.0048	14	0.752	0.127	9
Stegastes variabilis	Caribbean	Warm	0.134	0.012	33	0.025	0.005	20	0.626	0.136	11
		Cool	0.145	0.012	32	0.031	0.003	20	0.763	0.056	13
Stegastes acapulcoensis	Pacific	Warm	0.099	0.004	64	0.019	0.0029	31	0.448	0.065	22
		Cool	0.122	0.014	35	0.020	0.0014	6	-	-	-
Stegastes flavilatus	Pacific	Warm	0.080	0.004	65	0.016	0.0040	25	0.384	0.079	17
		Cool	0.101	0.011	75	0.020	0.0038	13	0.461	0.085	10

BLENNIIDAE

Ophioblennius macclurei	Caribbean	Warm	0.137	0.007	27	0.030	0.0027	27	0.728	0.062	12
		Cool	0.142	0.006	27	0.035	0.0013	16	0.859	0.041	16
Ophioblennius steindachneri	Pacific	Warm	0.110	0.008	37	0.025	0.0031	12	0.572	0.073	12
		Cool	0.134	0.013	46	0.031	0.0034	12	0.699	0.062	12

CHAENOPSIDAE

| *Acanthemblamaria rivasi* | Caribbean | Warm | 0.582 | 0.051 | 31 | 0.116 | 0.013 | 31 | 2.80 | 0.266 | 28 |

		Cool	0.584	0.036	9	0.138	0.012	8	3.20	0.289	8
Acanthemblamaria handcocki	Pacific	Warm	0.357	0.026	37	0.077	0.008	34	1.802	0.199	30
		Cool	0.344	0.018	13	0.088	0.007	12	2.034	0.160	12

GOBIIDAE

Coryphopterus dicrus	Caribbean	Warm	0.095	0.009	16	0.013	0.0014	14	0.297	0.020	11
		Cool	-	-	-	-	-	-	-	-	-
Coryphopterus glaucofrenum	Caribbean	Warm	0.077	0.011	42	0.009	0.0020	26	0.201	0.049	20
		Cool	0.097	0.033	23	0.011	0.0015	22	-	-	-
Coryphopterus personatus	Caribbean	Warm	0.072	0.007	28	0.010	0.0012	28	0.211	0.030	28
		Cool	-	-	-	-	-	-	-	-	-
Coryphopterus tortugae	Caribbean	Warm	0.106	0.009	29	0.011	0.0013	27	0.25	0.035	27
		Cool	-	-	-	-	-	-	-	-	-
Coryphopterus urospilus	Pacific	Warm	0.051	0.009	20	0.006	0.0004	14	0.178	0.031	30

Cool   0.078   0.027   33   0.008   0.0012   31   -   -   -

Supplementary Table 2: Regression equations for the relationship between egg volume (µl) and egg dry weight (µg) and energy content (joules) generated by ordinary least squares regression.

Species	Equation	r^2	n	p
POMACENTRIDAE				
Abudefduf saxatilis	Dry weight = 0.050 - 0.023*volume	0.55	66	ns
	Egg energy = 0.73 + 1.64*volume	0.06	48	ns
Abudefduf troschelii	Dry weight = 0.004 + 0.21*volume	0.21	54	<0.001
	Egg energy = 0.012 + 5.34*volume	0.24	44	<0.001
Chromis multilineata	Dry weight = 0.008 + 0.097*volume	0.14	15	ns
	Egg energy = -0.26 + 7.66*volume	0.81	6	0.01
Chromis atrilobata	Dry weight = 0.013 + 0.078*volume	0.04	15	ns
	Egg energy = 0.34 + 1.34*volume	0.01	14	ns
Microspathadon bairdii	Dry weight = 0.017 + 0.070*volume	0.04	16	ns
	Egg energy = 0.68 - 0.076*volume	0.001	10	ns
Microspathodon chrysurus	Dry weight = 0.013 + 0.11*volume	0.06	52	0.07
	Egg energy = 0.13 + 4.11*volume	0.15	38	0.02
Microspathodon dorsalis	Dry weight = 0.0063 + 0.15*volume	0.54	32	<0.001
	Egg energy = 0.19 + 3.20*volume	0.56	24	<0.001
Stegastes adustus	Dry weight = 0.0041 + 0.16*volume	0.16	49	0.004
	Egg energy = -0.0092 + 5.01*volume	0.29	40	<0.001
Stegastes diencaeus	Dry weight = -0.0098 + 0.28*volume	0.31	30	0.002
	Egg energy = 0.75 - 0.24*volume	0.0005	18	ns
Stegastes leucostictus	Dry weight = 0.0093 + 0.16*volume	0.18	56	0.001
	Egg energy = 0.36 + 3.61 *volume	0.23	41	0.002
Stegastes partitus	Dry Weight = 0.0041+ 0.15*vol	0.22	51	<0.001
	Egg energy = -0.051 + 5.70*vol	0.45	40	<0.001
Stegastes planifrons	Dry Weight = -0.0064 + 0.24*vol	0.28	35	0.001
	Egg energy = -0.29 + 6.92*vol	0.19	19	0.06
Stegastes variabilis	Dry weight = -0.014 + 0.28*vol	0.34	40	<0.001
	Egg energy = -0.21 + 6.13*vol	0.27	24	0.009

Stegastes acapulcoensis	Dry weight = 0.010 + 0.085*volume	0.05	37	ns
	Egg energy = -0.17 + 6.32*volume	0.10	22	ns
Stegastes flavilatus	Dry weight = 0.0014 + 0.19*volume	0.29	38	<0.001
	Egg energy = 0.10 + 3.68*volume	0.26	27	0.006

BLENNIIDAE

Ophioblennius macclurei	Dry weight = 0.0028 + 0.21*volume	0.22	43	0.001
	Egg energy =-0.43 + 8.79*volume	0.80	28	<0.001
Ophioblenniu steindachneri	Dry weight = 0.0099 + 0.16*volume	0.12	24	0.09
	Egg energy = 0.26 + 3.20*volume	0.12	24	ns

CHAENOPSIDAE

Acanthemblemaria rivasi	Dry weight = 0.017 + 0.18*volume	0.31	39	<0.001
	Egg energy = 0.92 + 3.35*volume	0.22	36	<0.01
Acanthemblemaria hancocki	Dry weight = 0.050 + 0.083*volume	0.05	46	ns
	Egg energy = 1.36 + 1.43*volume	0.03	42	ns

GOBIIDAE

Coryphopterus dicrus	Dry weight = 0.018 - 0.047*volume	0.15	11	ns
	Egg energy = 0.30 - 0.037*volume	0.00	11	ns
Coryphopterus glaucofrenum	Dry weight = 0.0068 + 0.035*volume	0.22	48	<0.001
	Egg energy = -0.044 + 3.22*volume	0.29	20	0.015
Coryphopterus personatus	Dry weight = 0.0043 + 0.072*volume	0.14	28	0.05
	Egg energy = 0.078 + 1.86*volume	0.17	28	0.03
Coryphopterus tortugae	Dry weight = 0.013 - 0.013*volume	0.009	27	ns
	Egg energy = 0.303 - 0.48*volume	0.02	27	ns
Coryphopterus urospilus	Dry weight = 0.0055 + 0.026*volume	0.28	45	<0.001
	Egg energy = 0.16 + 0.27*volume	0.06	30	ns

Supplementary Table 3: 2-way ANOVA analyses of log-transformed data for egg volume (µl), egg dry weight (µg) and egg energy content (j) of geminate pairs or geminate clades for each genus. Statistically significant p-values are highlighted in bold.

	df	F	p
POMACENTRIDAE			
Abudefduf			
Egg Volume			
Season	1	39.05	**<0.0001**
Species	1	1177.13	**<0.0001**
Species X Season	1	0.005	0.94
Egg Dry Weight			
Season	1	8.46	**0.004**
Species	1	223.84	**<0.0001**
Species X Season	1	4.71	**0.03**
Egg Energy Content			
Season	1	12.00	**0.0008**
Species	1	162.38	**<0.0001**
Species X Season	1	2.22	0.14
Chromis			
Egg Volume - eggs not comparable			
Egg Dry Weight			
Season	1	0.03	0.59
Species	1	23.39	**<0.0001**
Species X Season	1	0.004	0.95
Egg Energy Content - Insufficient data			
Microspathodon			
Egg Volume			
Season	1	55.51	**<0.0001**
Species	2	36.39	**<0.0001**
Species X Season	2	7.89	**0.0004**
Egg Dry Weight			
Season	1	6.07	**0.02**
Species	2	0.13	0.87
Species X Season	2	4.31	**0.02**
Egg Energy Content (w/o *M. bairdi*)			
Season	1	18.77	**<0.0001**
Species	1	0.57	0.45
Species X Season	1	6.15	**0.02**
Stegastes **clade III**			
Egg Volume			

Season	1	110.31	**<0.0001**
Species	1	296.46	**<0.0001**
Species X Season	1	15.22	**0.0002**
Egg Dry Weight			
Season	1	3.24	0.076
Species	1	66.33	**<0.0001**
Species X Season	1	0.31	0.58
Egg Energy Content - Insufficient data			

Stegastes - clade IV

Egg Volume

Season	1	201.13	**<0.0001**
Species	3	1326.14	**<0.0001**
Species X Season	3	39.23	**<0.0001**
Egg Dry Weight			
Season	1	6.77	**0.01**
Species	3	35.41	**<0.0001**
Species X Season	3	1.81	0.15
Egg Energy Content			
Season	1	4.97	**0.03**
Species	3	34.09	**<0.0001**
Species X Season	3	1.46	0.23

BLENNIIDAE

Ophioblennius

Egg Volume

Season	1	76.16	**<0.0001**
Species	1	116.80	**<0.0001**
Species X Season	1	37.26	**<0.0001**
Egg Dry Weight			
Season	1	56.41	**<0.0001**
Species	1	40.75	**<0.0001**
Species X Season	1	2.32	0.13
Egg Energy Content			
Species	1	49.53	**<0.0001**
Season	1	72.79	**<0.0001**
Species X Season	1	0.51	0.48

CHAENOPSIDAE

Acanthemblemaria

Egg Volume

Season	1	0.59	0.45
Species	1	704.79	**<0.0001**

Species X Season	1	1.30	0.26
Egg Dry Weight			
Season	1	29.84	**<0.0001**
Species	1	240.66	**<0.0001**
Species X Season	1	0.50	0.48
Egg Energy Content			
Season	1	23.72	**<0.0001**
Species	1	287.34	**<0.0001**
Species X Season	1	0.02	0.88

GOBIIDAE

Coryphopterus glaucofrenum vs. *C. urospilus*			
Egg Volume			
Season	1	41.09	**<0.0001**
Species	1	46.65	**<0.0001**
Species X Season	1	5.11	**0.03**
Egg Dry Weight			
Season	1	25.19	**<0.0001**
Species	1	72.57	**<0.0001**
Species X Season	1	1.63	0.21
Egg Energy Content - Insufficient data			